



Three new species from the subfamily Phyllocoptinae (Acari, Trombidiformes, Eriophyidae) in Iran

Parisa Lotfollahi¹, Enrico de Lillo², Karim Haddad Irani-Nejad³

1 Department of Plant Protection, Faculty of Agriculture, Azarbaijan Shahid Madani University, Tabriz, Iran 2 Department of Soil, Plant and Food Sciences (Di.S.S.P.A.), Entomology and Zoology Section, University of Bari Aldo Moro, via Amendola, 165/a, 70126 Bari, Italy 3 Department of Plant Protection, Faculty of Agriculture, University of Tabriz, Tabriz, Iran

Corresponding author: Enrico de Lillo (enrico.delillo@uniba.it)

Academic editor: Vladimir Pesic | Received 10 June 2014 | Accepted 2 July 2014 | Published 17 July 2014

http://zoobank.org/0825F733-2DFF-460E-9B7A-AC359960CE6E

Citation: Lotfollahi P, de Lillo E, Irani-Nejad KH (2014) Three new species from the subfamily Phyllocoptinae (Acari, Trombidiformes, Eriophyidae) in Iran. ZooKeys 426: 17–27. doi: 10.3897/zookeys.426.8087

Abstract

Three new eriophyid species (Phyllocoptinae), *Shevtchenkella denticulata* **sp. n.**, *Notallus pestehae* **sp. n.** and *Echinacrus ruthenicus* **sp. n.**, were described from *Eryngium thyrsoideum* Boiss. (Apiaceae), *Pistacia vera* L. (Anacardiaceae) and *Lycium ruthenicum* Murray (Solanaceae), respectively. All the three new species were collected from southwest of the East Azerbaijan province, Iran in 2011. It is the first record of an eriophyoid mite collected from *E. thyrsoideum* and *L. ruthenicum* and the first record of *Notallus* from Anacardiaceae plant family.

Keywords

East Azerbaijan, Notallus, Echinacrus, Shevtchenkella, Eriophyoidea, washing method

Introduction

As far as known concerning Iranian fauna, no eriophyoid species has been recorded from Apiaceae. Four eriophyoid species (*Aceria mangiferae* Sayed, 1946, *Aceria pistaciae* (Nalepa, 1899), *Aceria stefanii* (Nalepa, 1898) and *Calacarus citrifolii* Keifer, 1955) have been recorded from Anacardiaceae (Mehrnegad and Daneshvar 1991, Arbabi et al. 1999,

Mehrnejad and Ueckermann 2001, Khanjani and Haddad 2006), and five eriophyoid species [*Tetra lycopersici* Xue & Hong, 2005, *Aceria eucricotes* (Nalepa, 1892), *Aceria melongena* (Zaher & Abou-Awad, 1979), *Aculops lycopersici* (Tryon, 1917) and *Aculus solani* Boczek & Davis, 1984] have been recorded from Solanaceae (Sepasgozarian 1973, Ramazani et al. 2006, Xue et al. 2011, Gharezadeh et al. 2013).

Considering the relevance of this subject and the scientific importance of the evaluation of the mite fauna in scarcely known areas (de Lillo and Skoracka 2010), samples of *Eryngium thyrsoideum* Boiss. (Apiaceae), *Pistacia vera* L. (Anacardiaceae) and *Lycium ruthenicum* Murray (Solanaceae) plants were collected in Iran and their associated eriophyoid mites were studied.

Material and methods

The eriophyoid mite fauna of *E. thyrsoideum*, *P. vera* and *L. ruthenicum* was surveyed in the southwest of East Azerbaijan, Iran, during 2011. Mites were recovered from plant materials according to the modified washing method based on the protocol developed by Monfreda et al. (2007) and mounted on slides according to the protocol reported in Baker et al. (1996). The terminology and setal notation in the morphological descriptions follow mainly Lindquist (1996). The number of measured specimens (n) is given within parentheses in the description. All measurements were made with a phase contrast microscope Olympus BX50 according to Amrine and Manson (1996) and de Lillo et al. (2010), and are given in micrometres. Measurements and means are rounded off to the nearest integer when required, and refer to the length of the morphological characters unless specified otherwise. Since some measurements of the holotype could not be taken, due to the mounting position, the mean measurements of the paratypes are reported. Range values are given in parentheses except in case of constant value or unless specified otherwise. Drawings were made according to de Lillo et al. (2010) and abbreviations follow Amrine et al. (2003). The genus classification follows Amrine et al. (2003) and comparisons were also made with the new genera described since that publication.

Type materials are deposited in the collection of the Acarology Laboratory, Department of Plant Protection, Faculty of Agriculture, University of Tabriz, Tabriz (Iran) and of the Department of Soil, Plant and Food Sciences (Di.S.S.P.A.), section of Entomology and Zoology, University of Bari Aldo Moro (Italy).

Shevtchenkella denticulata sp. n.

http://zoobank.org/5118BBE1-2C15-4BD4-9DCB-F0E129215103 Fig. 1

Description. FEMALE. Body dorso-ventrally depressed, 205 (186–226, n = 10), 38 thick, 71 (68–77) wide. **Gnathosoma** 35 (31–38) projecting obliquely downwards, chelicerae 23 (23–32), setae *d* 6 (5–7) and unbranched. **Prodorsal shield** 44 (44–52)

included the frontal lobe, 73 (68–77) wide, semicircular in anterior shape with a broad, semicircular frontal lobe, 13 (12–16), over gnathosomal base provided with a spine on the lateral view. Shield pattern distinct and including 26 depressed cells; tubercles of setae sc on the rear shield margin 32 (32–37) apart, setae sc 8 (7–9), projecting posteriorly. Leg I 35 (32–37), femur 10, genu 4 (4–5), tibia 9 (8–10), tarsus 8 (7–8), ω 7 (6-7) and knobbed, empodium simple, 4 (4-4.5), 4-rayed; setae bv 10 (9-11), setae l'' 17 (15–20), setae l' 4 (4–5), setae ft' 17 (15–20), setae ft'' 20 (18–22). Leg II 32 (30-34), femur 10 (9-10), genu 4 (4-5), tibia 7 (6-7), tarsus 7 (7-8), ω 6 (6-7) and knobbed, empodium simple, 4, 4-rayed; setae by 10 (8–12), setae l'' 5 (5–7), setae ft'5 (4–5), setae ft'' 17 (17–20). **Coxae** with microgranules sometimes lined; setae 1b 8 (7-10), tubercles 1b 12 (11-19) apart, setae 1a 27 (26-31), tubercles 1a 8 (8-9) apart, setae 2a 45 (44–53), tubercles 2a 26 (23–26) apart. Prosternal apodeme 9 (8–10). **Opisthosoma** dorsally flat, with a large furrow and small lobes, 21 (21–24) broad and smooth dorsal semiannuli with the exception of the last two provided with spiny microtubercles protruding from the posterior margin of the annuli; 67 (67-81) narrow microtuberculated ventral semiannuli (counted since the first annulus after the coxae II); 9 (9–13) semiannuli between coxae and genital area plus 4–5 transversal rows of lined granules at the base of the genital coverflap. Small and circular microtubercles, closer to the posterior part of ventral semiannuli. Setae c2 25 (20-26) on ventral semiannulus 13 (12–17), setae d 59 (59–70) on ventral semiannulus 27 (27–35); setae e 15 (14-16) on ventral semiannulus 44(44-57); setae f28(26-30) on ventral semiannulus 63 (63–77). Last 4 ventral semiannuli with elongated linear microtubercles protruding from the posterior margin of the annuli. Setae h2 62 (62–78) very thin at the apex, h1 1-2. Genital coverflap 15 (13-18), 23 (23-27) wide, with 14 (13-15) striae and denticulate margin; setae 3a 20 (15–20) apart, 15 (14–17).

MALE. Similar in shape and prodorsal shield arrangement to female, 192 (n = 1). Prodorsal shield 48; setae sc 9, 34 apart; opisthosoma with 21 dorsal semiannuli and 68 ventral semiannuli; male genitalia 20 wide.

Type host plant. Eryngium thyrsoideum Boiss. (Apiaceae), Eringo or Sea Holly. Relation to the host plant. Vagrant on leaves; no apparent damage was observed. Type locality. Amir dizaj village, Azarshahr, Iran (37°40'17"N, 46°01'58"E), 1,950 m above sea level; late July 2011, coll. P. Lotfollahi.

Type material. Holotype: single female on a microscope slide (ET-IEA-AJ11L-1) (deposited at the Acarology Laboratory, Department of Plant Protection, Faculty of Agriculture, University of Tabriz, Tabriz, Iran). Paratypes: 12 females, 1 male and 2 nymphs mounted on separate microscope slides.

Other material. Mites preserved in Oudemans' fluid and extracted from the sample collected in the same locality on the same date above mentioned.

Etymology. This species is named based on the denticulate shape of the female genital coverflap.

Remarks. This is the first record of a species belonging to the genus *Shevtchenkella* collected on a plant of the family Apiaceae and the first record of an eriophyoid mite on *E. thyrsoideum*.

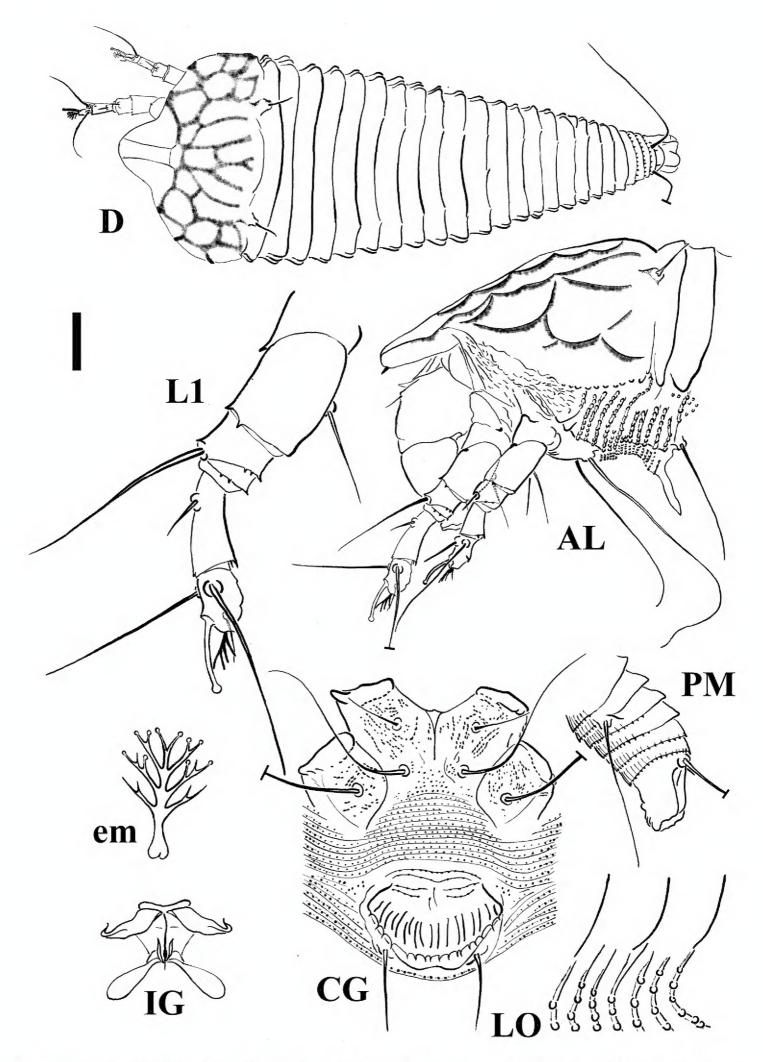


Figure 1. Schematic drawings of *Shevtchenkella denticulata* sp. n.: **AL** Lateral view of anterior body region **CG** Female coxigenital region **D** Dorsal view **em** Empodium **IG** Internal female genitalia **LO** Lateral view of annuli **L1** Leg I **PM** Lateral view of posterior opisthosoma. Scale bar: 17.5 μm for **D**; 10 μm for **AL**, **CG**, **IG**, **PM**; 5 μm for **LO**, **L1**; 2.5 μm for **em**.

Differential diagnosis. The new species herein described does not show any similarity with any known *Shevtchenkella* spp. whereas shows some similarities with *Aculus pimpinellae* (Liro, 1941) collected from *Pimpinella saxifraga* L. (Apiaceae) in Hollola, Hatsina, Tavastia australis Natural Province, Finland. Differences between these two species, other than those related to the fact they belong to two different genera, are: the ratio between the prodorsal shield length and the length of *sc* setae (5.5 in Iranian species *versus* 2 in Liro's species); number of dorsal annuli (21–24 in Iranian species *versus* 28 in Liro's species); size and shape of the female genital coverflap (15×23 with denticulate rear margin in Iranian species *versus* 15×16 with smooth margin in Liro's species).

Echinacrus ruthenicus sp. n.

http://zoobank.org/C54984F9-3B58-4756-A422-81683CE5C3A2 Fig. 2

Description. FEMALE (n=10). Body spindle shaped, 195 (195–255, including gnathosoma), 73 thick, 68 (68–79) wide. **Gnathosoma** 26 (25–37) projecting obliquely downwards, chelicerae 26 (22–30), setae d 7 (7–9), unbranched. **Prodorsal shield** 47 (47–54) included the frontal lobe, 70 (60–74) wide, sub-triangular with a broad based and distally pointed frontal lobe, 10 (8–11) over gnathosomal base (starting from the distal motivator end). Shield pattern reticulated, composed of 22 cells resulted of connecting distinct median, admedian, submedian and lateral lines with transverse lines. Tubercles of setae sc on the rear shield margin, 33 (28–35) apart, setae sc 16 (15–19), directing backward. Leg I 37 (35–38), femur 11 (10–12), genu 6 (5–6), tibia 10 (8– 10), tarsus 9 (8–9), ω 6.5 (6–7) distally knobbed, empodium simple, 4 (4–5), 4-rayed, rays distally funnel shaped; setae bv 13 (11–15), setae l'' 24 (22–26), setae l' 4 (3–5), setae ft' 20 (19–20), setae ft" 22 (22–23). Leg II 36 (32–36), femur 11 (10–11), genu 5 (5–6), tibia 8 (7–8), tarsus 8 (8–9), ω 6.5 (6–7) distally knobbed, empodium simple, 4 (4–5), 4-rayed; setae by 10 (9–11), setae l'' 5 (4–7), setae ft' 4, setae ft'' 21 (19–22). Coxae with lined dashes; setae 1b 7 (5-8), tubercles 1b 10 (9-12) apart, setae 1a 38 (27–38), tubercles 1a 7 (7–8) apart, setae 2a 60 (60–73), tubercles 2a 21 (21–26) apart. Prosternal apodeme 5 (5–6). **Opisthosoma** dorsally arched, with 44 (41–49) broad dorsal semiannuli, 76 (70-86) narrow ventral semiannuli (counted from the first annulus after the coxae II) and 11 semiannuli between coxae and genital coverflap plus 2–3 broken transversal rows of lined granules at the base of the coverflap. Triangular broad based microtubercles on the posterior margin of dorsal semiannuli with a lined longitudinal distribution; circular microtubercles, finely spiny, on the middle of ventral semiannuli; last 6 ventral semiannuli with elongated and linear microtubercles. Setae *c2* 45 (36–45) on ventral semiannulus 15 (12–17), setae *d* 70 (65–85) on ventral semiannulus 29 (25–34); setae *e* 58 (43–64) on ventral semiannulus 49 (44–57); setae f 29 (24–33) on ventral semiannulus 70 (64–80). 6 annuli after setae f. Setae h2 102 (92-112) very thin at the apex, $h1\ 2\ (2-3)$. Genital coverflap $14\ (11-16)$, $22\ (20-25)$ wide, with 12 (11–13) striae; setae 3a 18 (18–23), 15 (15–17) apart.

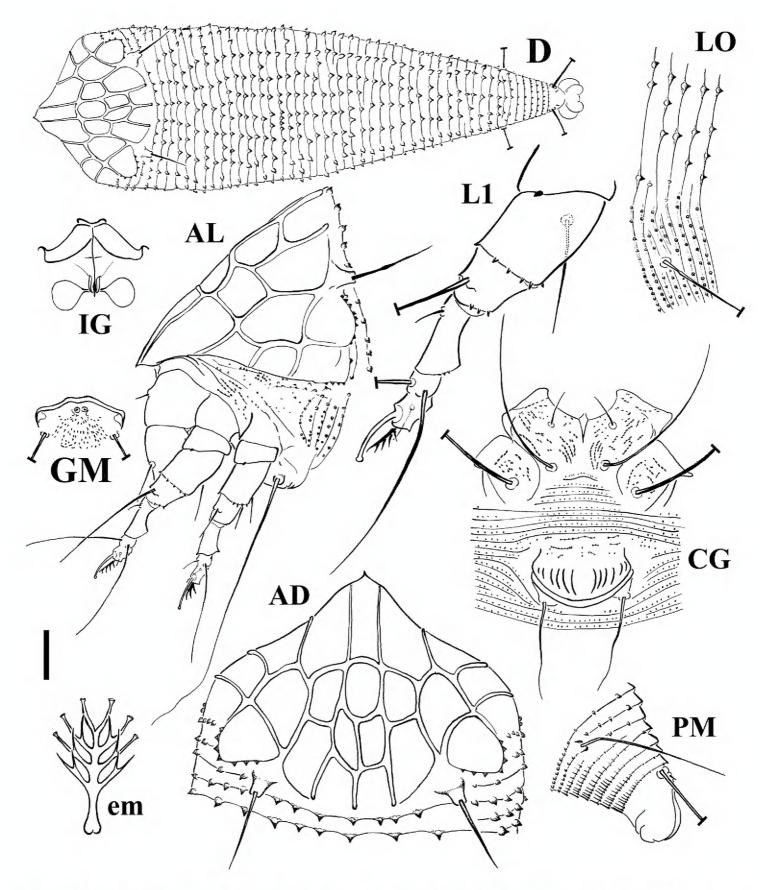


Figure 2. Schematic drawings of *Echinacrus ruthenicus* sp. n.: **AD** Dorsal view of anterior body region **AL** Lateral view of anterior body region **CG** Female coxigenital region **D** Dorsal view **em** Empodium **GM** Male genital region **IG** Internal female genitalia **LO** Lateral view of annuli **L1** Leg I **PM** Lateral view of posterior opisthosoma. Scale bar: 20 μm for **D**; 10 μm for **AD**, **AL**, **CG**, **IG**, **GM**, **PM**; 5 μm for **LO**, **L1**; 2.5 μm for **em**.

MALE (n=2). Similar in shape and prodorsal shield arrangement to female, 170–205. **Prodorsal shield** 45–50; setae *sc* 13–14, 23–32 apart. **Opisthosoma** with 39–44 dorsal semiannuli and 56–69 ventral semiannuli.

Type host plant. Lycium ruthenicum Murray (Solanaceae), Russian Box Thorn.

Relation to the host plant. Vagrant on leaves; no apparent damage was observed. **Type locality.** Ilkhchi, Iran (37°57′02″N, 45°58′40″E), 1,300 m above sea level; late July 2011, coll. P. Lotfollahi.

Type material. Holotype: single female on a microscope slide (LR-IEA-II11L-1) (at the Acarology Laboratory, Department of Plant Protection, Faculty of Agriculture, University of Tabriz, Tabriz, Iran). Paratypes: 9 females, 2 males and 1 nymph mounted on separate microscope slides.

Other material. Mites preserved in Oudemans' fluid as extracted from the same sample as the type specimens.

Etymology. The specific epithet is coming from the host plant name *ruthenicum*, deleting "m" and adding "s" as suffix.

Remarks. This is the first record of the genus *Echinacrus* on plants of family Solanaceae, first record of this genus in Iran and the first record of eriophyoid mites on *L. ruthenicum*.

Differential diagnosis. The new species herein described was compared with all *Echinacrus* species and similarities along with *E. septemcarinatus* (Liro, 1941), collected on *Frangula dodonei* Ard. (the synonym *Rhamnus frangula* L. was originally listed by Liro) in Lintula, Isthmus karelicus, Finland, were observed. The empodial rays (4 of the Iranian species *versus* 5 of Liro's species), shape, number and density of dorsal microtubercles (denser and more numerous in the Iranian species than those of Liro's description) and prodorsal shield pattern (22 cells in the Iranian species *versus* a lower number of cells in part differently arranged) are the main differences between the two species.

Notallus pestehae sp. n. http://zoobank.org/D550E12F-7D51-4AFA-AD2A-B5945717350D Fig. 3

Description. FEMALE (n=11). Body spindle shaped, 165 (156–185, including gnathosoma), 53 (48–57) thick, 52 (49–52) wide. **Gnathosoma** 41 (38–43) projecting obliquely downwards, chelicerae 37 (35–41), setae *d* 5 (4–5), unbranched. **Prodorsal shield** 39 (38–44) included the frontal lobe, 50 (46–50) wide, broad oval, with a broad based and distally truncated frontal lobe, 8 (7–11) over gnathosomal base. Shield pattern composed of a faint short median line on posterior ½ of prodorsal shield, complete admedian lines close together in the middle of the prodorsal shield, and short first submedian lines on posterior 2/3 of the prodorsal shield, connected to admedian lines with a pair of transverse lines. Admedian lines delimit a median obscure strip (Fig. 3-AD). Tubercles of setae *sc* on the rear shield margin, 25 (24–26) apart, setae *sc* 42 (37–45), directing backward. **Leg I** 26 (25–28), femur 9 (7–9), genu 5 (4–5), tibia 5 (5–6), tarsus 6 (6–8), ω 7 (6.5–8) distally knobbed, empodium simple, 3.5 (3–4), 4-rayed; setae *bv* 11 (9–13), setae *l*" 19 (18–20), setae *l*' 7 (5–7), setae *ft*' 15 (12–16), setae *ft*" 17 (17–19). **Leg II** 20 (20–23), femur 7, genu 3 (3–4), tibia 4 (3–4), tarsus 6 (6–7), ω 7.5 (6.5–8) distally knobbed, empodium simple, 3.5 (3–4),

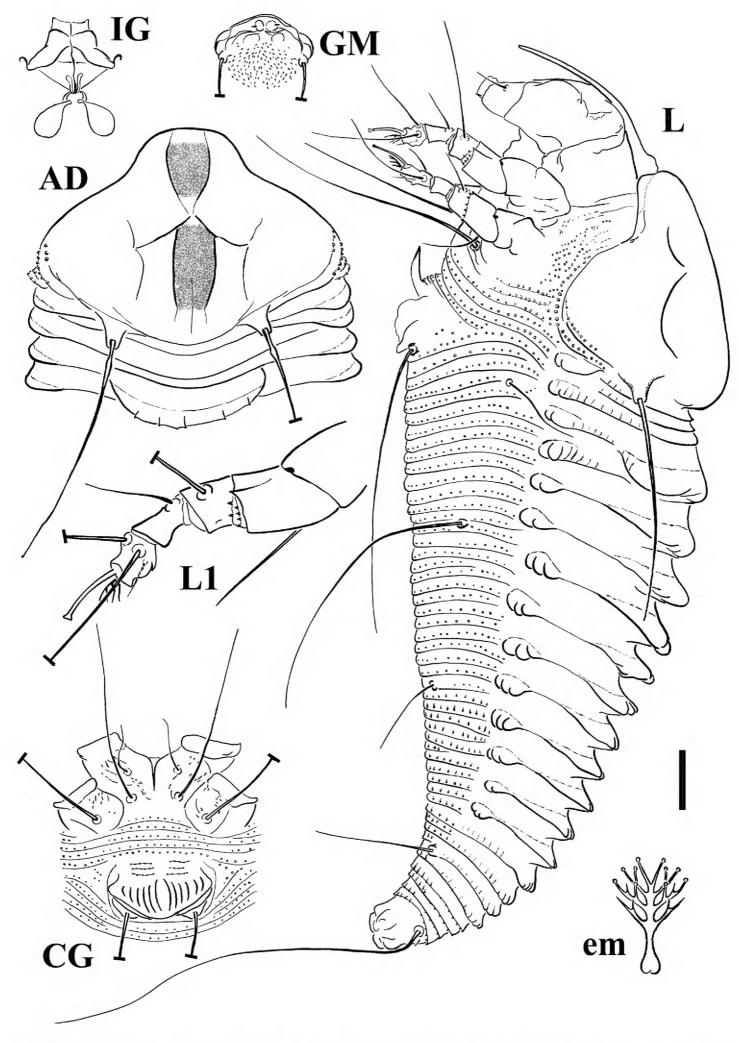


Figure 3. Schematic drawings of *Notallus pesthae* sp. n.: **AD** Dorsal view of anterior body region **CG** Female coxigenital region **em** Empodium **GM** Male genital region **IG** Internal female genitalia **L** Lateral view **L1** Leg I. Scale bar: 10 μm for **AD**, **CG**, **IG**, **GM**, **L**, **PM**; 5 μm for **L1**; 2.5 μm for **em**.

4-rayed; setae bv 11 (10–12), setae l'' 6 (6–7), setae ft' 6 (4–6), setae ft'' 16 (15–18). **Coxae** with sparse dashes in part lined; setae 1b 7 (7–9), tubercles 1b 8 apart, setae 1a 28 (27–33), tubercles 1a 6 (6–7) apart, setae 2a 45 (37–55), tubercles 2a 17 (17–18) apart. Prosternal apodeme 6 (6–6.5). **Opisthosoma** with 22 (21–23) broad dorsal semiannulis provided with three dorsal ridges; median ridge from forth dorsal semiannulus extended up to 16 (16–17) semiannulus, lateral ridges from first dorsal semiannulus extended up to 16 semiannulus; faint elongated microtubercles on the ridges; 59 (53–59) narrow microtuberculated ventral semiannuli (counted from the first annulus after the coxae II) and 5 semiannuli between coxae and genital coverflap plus 3 transversal rows of lined granules at the base of the coverflap. Setae c2 13 (11–15) on ventral semiannulus 11 (9–11), setae d 50 (43–51) on ventral semiannulus 22 (20–22); setae e 13 (13–15) on ventral semiannulus 39 (33–39); setae f 20 (15–23) on ventral semiannulus 54 (48–54). 5 annuli after setae f. Setae h2 53 (40–70) very thin at the apex, h1 very minute about 1. **Genital coverflap** 8 (8–11), 18 (18–19) wide, with 14 (12–14) striae; setae f3 (43–52), 11 (10–13) apart.

MALE (n=2). Similar in shape and prodorsal shield arrangement to female, 160–168. **Prodorsal shield** 37–41; setae *sc* 24–31, 23 apart. **Opisthosoma** with 22 dorsal semiannuli and 49–51 ventral semiannuli; genital region 17 wide; setae *3a* 41.

Type host plant. Pistacia vera L. (Anacardiaceae), Pistachio.

Relation to the host plant. Vagrant on leaves; no apparent damage was observed. **Type locality.** Akhijahan village, Gogan, Iran (37°47'14"N, 45°57'03"E), 1,346 m above sea level; late July 2011, coll. P. Lotfollahi.

Type material. Holotype: single female on a microscope slide (PV-IEA-AN11L-1) (deposited at the Acarology Laboratory, Department of Plant Protection, Faculty of Agriculture, University of Tabriz, Tabriz, Iran). Paratypes: 11 females and 4 males mounted on separate microscope slides.

Other material. Mites preserved in Oudemans' fluid as extracted from the same sample as the type specimens.

Etymology. The specific epithet is coming from the Persian common name *pesteh* given to pistachio.

Remarks. This is the first record of a species belonging to the genus *Notallus* on plants of the Anacardiaceae family.

Differential diagnosis. The genus *Notallus* is characterized by both lateral and middorsal ridges beginning on the forth dorsal semiannulus (Amrine et al. 2003) while the Iranian mite is provided with lateral ridges beginning since the first dorsal semiannulus. In addition, *N. nerii* Keifer, 1975 has more dorsal semiannuli (about 26) and less ventral semiannuli (about 49) in respect to *N. pestheae* (about 22 and 59, respectively), its prodorsal shield is provided with a narrower frontal lobe and an almost "obsolete" pattern composed of faint admedian and converging submedian lines (*N. pestheae* displays a clear pattern). Finally, *N. pterocaryae* Kuang, Luo & Wang, 2005, has smooth prodorsal shield and coxae (both areas are provided with ornamentations in *N. pestheae*) and empodium 7-rayed (4-rayed in *N. pestheae*).

Acknowledgements

The authors are grateful to Prof. Radmila Petanović (Department of Entomology and Agricultural Zoology, Faculty of Agriculture, University of Belgrade, Serbia) for her critical review of this manuscript. This research was partially supported by the University of Tabriz, Iran, and University of Bari Aldo Moro, Italy.

References

- Amrine JW Jr, Manson DCM (1996) Preparation, mounting and descriptive study of Eriophyoid mites. In: Lindquist EE, Sabelis MW, Bruin J (Eds) Eriophyoid Mites. Their Biology, Natural Enemies and Control. World Crop Pests, 6, Elsevier Science Publishers, Amsterdam, Netherlands, 383–396. doi: 10.1016/S1572-4379(96)80023-6
- Amrine JW Jr, Stasny TA, Flechtmann CHW (2003) Revised keys to world genera of Eriophyoidea (Acari: Prostigmata). Indira Publishing House, West Bloomfield, Michigan, 244 pp.
- Arbabi M, Kamali H, Mohsenin AB, Baradaran P (1999) Eriophyid mites status on fruit trees of Iran. Acarological Society of India, Bangalore, Symposium, 27–30 Oct.
- Baker EW, Kono T, Amrine JW Jr, Delfinado-Baker M, Stasny TA (1996) Eriophyoid mites of the United States. Indira Publishing House, West Bloomfield, Michigan, USA, 394 pp.
- Boczek J, Davis R (1984) New species of eriophyid mites (Acari: Eriophyoidea). Florida Entomologist 67(2): 198–213. doi: 10.2307/3493939
- de Lillo E, Skoracka A (2010) What's "cool" on Eriophyoid Mites? Experimental and Applied Acarology 51(1–3): 3–30. doi: 10.1007/s10493-009-9297-4
- de Lillo E, Craemer C, Amrine JW Jr, Nuzzaci G (2010) Recommended procedures and techniques for morphological studies of Eriophyoidea (Acari: Prostigmata). Experimental and Applied Acarology 51(1–3): 283–307. doi: 10.1007/s10493-009-9311-x
- Gharezadeh M, Kamali H, Shirdel D (2013) Mite fauna of the superfamily Eriophyoidea (Acari: Prostigmata) associated with landscape plants and trees in Mashhad city, Iran. The 2nd International Persian Congress of Acarology, 29–31 August, 2013: 68.
- Keifer HH (1955) Eriophyid Studies XXIII. Bulletin of the Department of Agriculture, State of California 44: 126–130.
- Keifer HH (1975) Eriophyid Studies C-10. Agricultural Research Service. United States Department of Agriculture, 1–24.
- Khanjani M, Haddad-Iraninejad K (2006) Injurious Mites of Agricultural Crops in Iran. Bu-Ali Sina University of Hamadan Press, 515 pp.
- Kuang H-Y, Luo G-H, Wang A-W (2005) Fauna of Eriophyid Mites from China (II) (Acari: Eriophyoidea). China Forestry Publ. House, Beijing, 176 pp.
- Lindquist EE (1996) External anatomy and notation of structures. In: Lindquist EE, Sabelis MW, Bruin J (Eds) Eriophyoid Mites. Their Biology, Natural Enemies and Control. World Crop Pests, 6, Elsevier Science Publishers, Amsterdam, Netherlands, 3–31. doi: 10.1016/S1572-4379(96)80003-0

- Liro JI (1941) Über neue und seltene Eriophyiden (Acarina). Annales Zoologici Societatis Zoologicae-Botanicae Fennicae, Vanamo 8(7): 1–53.
- Mehrnegad MR, Daneshvar H (1991) First report of two eriophyid mites from pistachio in Kerman and Yazd. Applied Entomology and Phytopathology 58(1–2): 55.
- Mehrnejad MR, Ueckermann EA (2001) Mites (Arthropoda, Acari) associated with pistachio trees (Anacardiaceae) in Iran (I). Systematic and Applied Acarology, Special Publication 6: 1–12.
- Monfreda R, Nuzzaci G, de Lillo E (2007) Detection, extraction, and collection of Eriophyoid mites. Zootaxa 1662: 35–43.
- Nalepa A (1892) Neue Gallmilben. 4. Fortzung. Anzeiger der kaiserlichen Akademie Wissenschaften. Mathematische–naturwissenschaftliche Klasse, Wien 29(13): 128.
- Nalepa A (1898) Neue Gallmilben. 16. Fortzung. Anzeiger der kaiserlichen Akademie Wissenschaften. Mathematische–naturwissenschaftliche Klasse, Wien 35(17): 163–164.
- Nalepa A (1899) Neue Gallmilben. 18. Fortzung. Anzeiger der kaiserlichen Akademie Wissenschaften. Mathematische–naturwissenschaftliche Klasse, Wien 36(17): 217–218.
- Ramazani L, Mosaddegh MS, Shisheh P, Kamali K (2006) Seven new records of eriophyoid mites on weeds from Iran. The Proceedings 17th Plant Protection Congress Iran, 185 pp.
- Sayed MT (1946) Aceria mangiferae nov. spec. (Eriophyes mangifeare Hassan MS) (Acarina-Eriophyidae). Bulletin de la Société Fouad Ier d'Entomologie 30: 7–10.
- Sepasgozarian H (1973) Mites and their economic important in Iran. Proceedings of the 3rd International Congress on Acarology, Dr. Junk, Publ., The Hague Academia, 1971, 241–245.
- Tryon H (1917) Report of the Entomologist and Vegetable Pathologist. Queensland Department of Agriculture & Stork Report 1916/1917: 49–63.
- Xue X-F, Hong X-Y (2005) Five new species of the genus *Tetra* Keifer (Acari: Eriophyoidea) from China. Zootaxa 1067: 37–48.
- Xue X-F, Sadeghi H, Hong H-Y, Sinaie S (2011) Nine eriophyoid mite species from Iran (Acari, Eriophyidae). ZooKeys 143: 23–45. doi: 10.3897/zookeys.143.2162
- Zaher MA, Abou-Awad BA (1979) Three new species of the genera *Eriophyes* and *Phytoptus* in Egypt. (Eriophyoidea: Eriophyidae). Acarologia 20(4): 556–562.